

Asian Pacific Journal of Reproduction

Offiial publication of Hainan Medical University



# **Editorial Board**

**Editor-in-Chief** Yuan-hua Huang, Ph.D. Professor Hainan Medical University, Haikou, China

#### Executive Editors-in-Chief

Eric Chung Professor AndroUrology Centre for Sexual, Urinary and Reproductive Excellence; Department of Urology, Princess Alexandra Hospital, Brisbane, Australia

Bo Cui, M.D., Ph.D. Hainan Medical University, Haikou, China

#### Associate Editor-in-Chief

Gian Mario Tiboni, M.D. Associate Professor Obstetrics and Gynecology, Department of Medicine and Aging Sciences University"G.d'Annunzio" Chieti Pescara, Italy

#### **Editorial Director**

Jing Li Hainan Medical University, Haikou, China

#### Managing Editors

Yan Lei Hainan Medical University, Haikou, China

Luying Lin Hainan Medical University, Haikou, China

#### Editorial Board

Asmaa Gaber Abdou Professor Pathology Department, Faculty of Medicine, Menoufiya University, Egypt

Juan F. Aller, Ph.D. Researcher

National Institute of Agricultural Technology, Experimental Station of Balcarce, Department of Animal Production, Animal Reproductive Biotechnology, Buenos Aires Province, Argentina

Kristian Almstrup, Ph.D. Senior Scientist

Department of Growth and Reproduction, Copenhagen University Hospital, Rigshospitalet, Section GR-5064, 9 Blegdamsvej, DK-2100 Copenhagen, Denmark

Brent M. Bany, Ph.D. Associate Professor Department of Physiology, Southern Illinois University School of Medicine, Carbondale, Illinois, USA

Mostafa Borahay, Ph.D. Associate Professor Department of Gynecology and Obstetrics, Johns Hopkins School of Medicine, Johns Hopkins Bayview Medical Center, Baltimore, USA

Luciana Bordin, Ph.D. Department of Molecular Medicine-Biological Chemistry, University of Padova, Viale G. Colombo 3, 35131 Padua, Italy

Sandip Chattopadhyay Department of Biomedical Laboratory Science and Management, Vidyasagar University, Midnapore, West Bengal, India

Zi-Jiang Chen, Ph.D. Professor Center for Reproductive Medicine, Cheeloo College of Medicine, Shandong University, Jinan, China; The Key Laboratory of Reproductive Endocrinology of Ministry of Education, Jinan, China; National Research Center for Assisted Reproductive Technology and Reproductive Genetics, Jinan, China; Center for Reproductive Medicine, Ren Ji Hospital, Shanghai, China; School of Medicine, Shanghai Jiao Tong University, Shanghai, China; Shanghai Key Laboratory of Assisted Reproduction and Reproductive Genetics, Shanghai, China Daniel G. Cyr, Ph.D. Professor Canada Research Chair in Reproductive Toxicology, INRS-Institut Armand Frappier, Université du Québec, Canada Pedro Alejandro Orihuela Díaz, Ph.D. Associate Professor Laboratory of Immunology of Reproduction, Faculty of Chemistry and Biology, University of Santiago, Chile Jens Fedder, Ph.D. Professor Centre of Andrology & Fertility Clinic, Dept. D, Odense University Hospital, Odense, Denmark Pasquale De Franciscis, Ph.D. Professor Department of Woman, Child and of General and Specialized Surgery Largo Madonna Delle Grazie 180138 Napoli, Italy Nasser Ghanem, Ph.D. Associate Professor Reproductive Biology & Genomics, Animal Production Department, Faculty of Agriculture, Cairo University, Giza, Egypt Antonio Gonzalez-Bulnes, Ph.D. Professor Comparative Physiology Group SGIT-INIA, Avda. Puerta de Hierro s/n 28040-Madrid, Spain Guo-ning Huang, Ph.D. Professor Chongqing Key Laboratory of Human Embryo Engineering, Chongqing Reproductive and Genetics Institute, Chongqing Health Center for Women and Children, Chongqing, China Rafiqul Islam, Ph.D. Principal Scientist Animal Reproduction & Gynaecology, ICAR-National Organic Farming Research Institute, Tadong, Gangtok -737102, Sikkim, India Kampon Kaeoket, Ph.D. Associate Professor Department of Clinical Sciences and Public Health, Faculty of Veterinary Science, Mahidol University, Nakhon Pathom, Thailand Sae Chul Kim, Ph.D. Professor Director General, Myongji Medical Center, Goyang, Gyeonggi-do, Korea; Emeritus Professor, Department of Urology College of Medicine, Chung-Ang University, Seoul, Korea; Chairman, Korean Prostate Health Council, Korea; Life member, Korean Academy of Science and Technology, National Academy of Medicine of Korea, Korea Marcel Leppée, Ph.D. Senior Researcher Institute for Healthy Ageing, Zagreb, Croatia Da-jin Li, Ph.D. Professor

Lab for Reproductive Immunology, Institute of Obstetrics & Gynecology, Fudan University Shanghai Medical College, Shanghai, China Maria de Lourdes Gomes Pereira, Ph.D. Professor Department of Medical Sciences, University of Aveiro, Campus Santiago, 3810-193 Aveiro, Portugal Puttaswamy Manjunath, Ph.D. Professor Department of Medicine, Faculty of Medicine, University of Montreal, Montreal, Canada Luc J Martin, Ph.D. Associate Professor Department of Biology, Faculty of Science, University of Moncton, 18 Avenue Antonine Maillet, Moncton, (Nouveau-Brunswick), Canada Martín Martínez-Torres Laboratorio de Biología de la Reproducción-Unidad de Morfofisiología, Facultad de Estudios Superiores Iztacala, Universidad Nacional Autónoma de México, México Premendu Prakash Mathur, Ph.D. Professor Department of Biochemistry & Molecular Biology, School of Life Sciences, Pondicherry University, Pondicherry-605014, India Liselotte Mettler, Ph.D. Professor Department of Obstetrics and Gynecology, University Clinics of Schleswig Holstein, Kiel, Germany Hossein Mozdarani, Ph.D. Professor Department of Medical Genetics, Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran Guilherme de Paula Nogueira, Ph.D. Full Professor Animal Physiology at the Department of Basic Science, Production and Animal Health (DAPSA), at the Aracatuba Veterinary Faculty (FMVA), linked to the Sao Paulo State University (UNESP), Brazil Stefan du Plessis. Ph.D. Professor Division of Medical Physiology, Department of Biomedical Sciences, Faculty of Medicine and Health Sciences, Stellenbosch University, Tygerberg, South Africa Giulia Ricci, Ph.D. Aggregate Professor Department of Experimental Medicine, Università della Campania "Luigi Vanvitelli", Via Luigi de Crecchio 7, 80138-Naples, Italy Fang-xiong Shi, Ph.D. Professor Department of Animal Reproduction and Breeding, College of Animal Science and Technology, Nanjing Agricultural University, Weigan 1, Nanjing, 210095, China Ying-pu Sun, Ph.D. Professor Center for Reproductive Medicine, Henan Key Laboratory of Reproduction and Genetics, The First Affiliated Hospital of Zhengzhou University, Zhengzhou, Henan, 450052, China Gaffari Turk, Ph.D. Professor Department of Reproduction and Artificial Insemination, Faculty of Veterinary Medicine, Firat University, Elazig,

Turkey

Jun-qing Wu, Ph.D. Professor Department of Epidemiology and Social Science on Reproduction of Shanghai Institute of Planned Parenthood Research, Shanghai, China; Fudan University, Shanghai, China

# Table of Contents





Click here to view optimized website for mobile devices



# Effect of different concentration of fish oil in skim milk-egg yolk extenders on post- thawed semen qualities of Kalang swamp buffalo bull

<u>Abdul Malik</u><sup>1</sup>, <u>A Jaelani</u><sup>1</sup>, <u>Neni Widaningsih</u><sup>1</sup>, <u>Gt Khairun Ni'mah</u><sup>1</sup>, <u>Raviani</u><sup>2</sup>, <u>Sakiman</u><sup>3</sup>, <u>N Sasongko</u><sup>3</sup> <sup>1</sup> Department of Animal Science, Faculty of Agriculture, Islamic University of Kalimantan, Banjarmasin-South Kalimantan, Indonesia

<sup>2</sup> Office of Animal Husbandry District of Banjar, South Kalimantan Province, Indonesia

<sup>3</sup> Centre of Artifical Insemination, Banjarbaru, South Kalimantan Province, Indonesia

Date of Submission12-Feb-2018Date of Decision10-Mar-2018

Date of Acceptance 05-Apr-2018

Date of Web Publication 31-May-2018



#### Correspondence Address:

Abdul Malik Department of Animal Science, Faculty of Agriculture, Islamic University of Kalimantan, Banjarmasin-South Kalimantan Indonesia

🖄 Login to access the email ID

Source of Support: None, Conflict of Interest: None



DOI: 10.4103/2305-0500.233576

© Get Permissions for commercial use

## How to cite this article:

Malik A, Jaelani A, Widaningsih N, Ni'mah G, Raviani, Sakiman, Sasongko N. Effect of different concentration of fish oil in skim milk-egg yolk extenders on post- thawed semen qualities of Kalang swamp buffalo bull. Asian Pac J Reprod 2018;7:139-42

## How to cite this URL:

Malik A, Jaelani A, Widaningsih N, Ni'mah G, Raviani, Sakiman, Sasongko N. Effect of different concentration of fish oil in skim milk-egg yolk extenders on post- thawed semen qualities of Kalang swamp buffalo bull. Asian Pac J Reprod [serial online] 2018 [cited 2022 Apr 27];7:139-42. Available from: <a href="https://www.apjr.net/text.asp?2018/7/3/139/233576">https://www.apjr.net/text.asp?2018/7/3/139/233576</a>



Asian Pacific Journal of Reproduction



Journal homepage: www.apjr.net

doi: 10.4103/2305-0500.233576 ©2018 by the Asian Pacific Journal of Reproduction. All rights reserved.

# Effect of different concentration of fish oil in skim milk-egg yolk extenders on postthawed semen qualities of Kalang swamp buffalo bull

Abdul Malik<sup>1</sup>, Jaelani A<sup>1</sup>, Neni Widaningsih<sup>1</sup>, Gt Khairun Ni'mah<sup>1</sup>, Raviani<sup>2</sup>, Sakiman<sup>3</sup>, Sasongko N<sup>3</sup>

<sup>1</sup>Department of Animal Science, Faculty of Agriculture, Islamic University of Kalimantan, Banjarmasin–South Kalimantan, Indonesia <sup>2</sup>Office of Animal Husbandry District of Banjar, South Kalimantan Province, Indonesia <sup>3</sup>Centre of Artifical Insemination, Banjarbaru, South Kalimantan Province, Indonesia

#### ARTICLE INFO

Article history: Received 12 February 2018 Revision 10 March 2018 Accepted 5 April 2018 Available online 31 May 2018

Keywords: Swamp buffalo Fish oil Semen cryopreservation Viability Plasma membrane integrity

## ABSTRACT

**Objective:** To explore the effect of fish oil at different concentrations on post-thawed semen of Kalang swamp buffalo. **Methods:** A total of 4 Kalang swamp buffalo bulls with 3-5 years of age and weighed about 340-360 kg were slected. Semen was regularly collected from these buffalo bulls once a week by an artificial vagina. Fish oil was supplementary at the dosages of 0 mg (control), 50 mg, 100 mg, 150 mg, and 200 mg to the extender (skim milk-egg yolk). Fresh, pre-freezing and frozen semen were thawed at 37 °C and evaluated for motility, viability, morphology, and plasma integrity of membrane. **Results:** The study results indicated that before freezing, supplementation of fish oil at the dose of 150 mg in the extender had significantly motility. And a significant (P<0.05) increase was observed in viability and motility of post-thawed semen at the dose of 150 mg fish oil, which was in difference with other treatment groups. **Conclusions:** Addition of 150 mg fish oil in the extender could be positive for the enhancement of the quality of post-thawed semen of Kalang swamp buffaloes.

#### 1. Introduction

Kalang swamp buffalo is a species of swamp buffalo, a native from Amuntai district, South Kalimantan province, Indonesia. Its pattern of maintenance differs from other types of swamp buffalo in the world. Everyday early in the morning, the buffalo go out of the cage (Kalang) to soak in the swamp for a day and then afternoon climb into the cage for rest. The population of Kalang swamp buffalo during five years has decreased, it is due to the number of cuts that is unbalanced with the development of the population and the low fertility[1]. One of the important factors to increase kalang swamp buffalo population is a estrus synchronization program trailed by artificial insemination (AI), because Kalang buffalo has unclear symptoms of estrus. While, an significant factor that determines the success of AI is the qualities of post-thawing semen that will affect conception and percentage of pregnancy[2].

Application of AI program in swamp buffaloes has been reported, but the results are not as good as in the application of AI in cows. The main cause of low AI success in swamp buffaloes is due to low quality of post-thawed sperm and low fertility level of female buffalo[3–7]. Furthermore, Andrabi[8] reported that there were many aspects that could affect the sperm motility, integrity of membrane and sperm viability through storage including cryopreservation and thawing process.

<sup>&</sup>lt;sup>EE</sup>First and corresponding author: Abdul Malik, Department of Animal Science, Faculty of Agriculture, Islamic University of Kalimantan, Banjarmasin–South Kalimantan, Indonesia. E-mail: sidol\_99@yahoo.com

Foundation project: This study was supported by APBU Islamic University of Kalimantan, Banjarmasin, Indonesia.

This is an open access article distributed under the terms of the Creative Commons Attribution-Non Commercial-Share Alike 3.0 License, which allows others to remix, tweak and buid upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

<sup>©2018</sup> Asian Pacific Journal of Reproduction Produced by Wolters Kluwer- Medknow How to cite this article: Abdul Malik, Jaelani A, Neni Widaningsih, Gt Khairun Ni'mah, Raviani, Sakiman, et al. Effect of different concentration of fish oil in skim milk-egg yolk extenders on post-thawed semen qualities of Kalang swamp buffalo bull. *Asian Pac J Reprod* 2018; 7(3): 139-142.

140

The freezing process of semen needs a step-by-step process that begins with microscopic and macroscopic semen evaluation, dilution, cooling, equilibration and freezing weakens the naturally existing seminal antioxidant capacity. Consequently, many strategies have been employed to progress the efficiency of cryopreservation process. However, the overall success to increase the effectiveness of the cryopreservation of buffalo semen remains relative low. In this condition, the exogenous antioxidants addition in extender has provided a great opportunity to increase sperm quality to struggle the oxidative stress during cryopreservation<sup>[9]</sup>.

One of the ingredients that can be used to help increase the sperm quality of buffalo post-thawing is fish oil. Fish oil contains several active ingredients such as omega 3 which is the main source of eicosapentaenoic acid, docosahexaenoic acid, saturated fatty acid and polyunsaturated fatty acids (PUFA)[10,11]. Several researches have been done on the effects of using fish oil to increase the semen quality either through feed or mixed with diluents in domestic animals. Meanwhile, the use of fish oil containing saturated fatty acids has been stated to be used to increase spermatozoa survival in general, membrane integrity and motility in boar[12] and cattle[13,14].

However, the influence of fish oil on Kalang buffalo spermatozoa has never been conducted. Therefore, the objective of the present study was carried out to determine the influence of fish oil supplementation in the extender on the sperm motility, viability, abnormality and integrity of membrane of cryopreserved buffalo bull sperm.

#### 2. Materials and methods

The study was conducted at the center of artificial insemination, province of South Kalimantan at Banjarbaru, Indonesia. The approval of the animal ethics committee on the conduct of this study was not necessary because there no invasive techniques on animals. Semen was used in the process of collection, evaluation and freezing by using standard method under the central artificial insemination program.

A total of 4 Kalang swamp buffalo bulls aged 3-5 years and weighed about 340-360 kg were used in the study. All bulls were kept in individual cage with the same feed. Forage was given to livestock about 10% of body weight of buffalo bull. Concentrate containing approximately 16% crude protein and 2.6% crude fat was given 3 kg/buffalo/day to each buffalo bull and water ad libitum. Semen was regularly collected from all bulls once a week by an artificial vagina. A total of 24 samples were poised and then transferred at 37  $^{\circ}$ C in water bath to the center of artificial insemination of Banjarbaru for evaluation.

## 2.1. Semen handling

The extender base used comprises of skim milk and egg yolk,

containing 10% skim mlik, 5% (v/v) egg yolk, 1% (w/v) fructose (Scharlau, Barcelona, Spain), 8% (v/v) glycerol (Merck, Darmstadt, Germany), penicillin (1 000 IU/mL) and streptomycin (1 000 mg/mL). The samples were designed to compare the effect of different concentrations of fish oil (liquid) (Manufacture Pty, Ltd. Minna Close Belrose, Australia) with the dose of 0 mg (control), 50 mg, 100 mg, 150 mg or 200 mg in the 100 mL skim milk-egg yolk extender. Semen was diluted with final sperm concentration  $25 \times 10^6$ /mL. Diluted semen was gradually cooled to 4 °C for 2 h. After equilibration, cooled semen was wrapped in 0.5 mL French straws, then placed on top of liquid nitrogen vapor (about 5 cm above liquid nitrogen) for 10 min. The straws were then dipped continuously and stored into liquid nitrogen. Later one week of kept in liquid nitrogen, five semen straws from each treatment group in each replicate were melted at 37  $^\circ\!\!\mathbb{C}$  for 30 s and analyzed for the following parameters.

#### 2.2. Evaluation of sperm qualities

Motility, viability, and abnormality parameters was measured on fresh semen and post-thawing semen. Motility of sperm was evaluated when a small droplet (10  $\mu$ L) of fresh, pre-freezing sperm and post-thawing sperm was placed in the center of a pre-warmed slide and then covered with cover slip, and observed under phase contrast microscope at 400× magnification. Morphology of sperm and viability were observed by using eosin-nigrosin stain as adopted by Khumran *et al*[15]. At least of 200 sperms were counted in four microscopic diverse fields adopted by Memon *et al*[16]. Morpholog of sperm was observed using the same slide used for sperm viability. The normal of sperm cells was counted from a total sperm cells examined. The sperm integrity of membrane was assessed in fresh, pre-freezing semen, and post-thawed semen. Integrity of membrane was evaluated using hypo osmotic swelling test as defined by Kaka *at al*[17].

#### 2.3. Statistical analysis

The effects of difference concentration of fish oil in the extender on motility, viability, integrity of membrane, and abnormality were analyzed by one-way analysis of variance (ANOVA). The collected data were presented as mean  $\pm$ SD, and were significant at *P*<0.05.

#### 3. Results

Observation of fresh semen by macroscopic and microscopic evaluation showed the following results: the average volume was  $(2.90\pm0.09)$  mL, pH was  $6.80\pm0.15$  and semen concentration was  $1.5\times10^{6}$ /mL. The average percentages of fresh semen were  $(71.00\pm1.04)\%$ ,  $(78.00\pm0.81)\%$ ,  $(15.00\pm1.02)\%$  and  $(73.00\pm0.93)\%$  for sperm motility, viability, abnormality and integrity of membrane,

respectively. The sperm motility of Kalang swamp buffalo bull semen stored for 4 h before freezing at dose of 150 mg fish oil was higher (P<0.05) than those at dose of 50 mg and 100 mg fish oil; however, the treatment with 200 mg fish oil caused a significant (P<0.05) reduction in motility (Table 1).

Significant (P<0.05) increases were detected in semen motility and viability of post frozen-thawed sperm at dose of 150 mg fish oil added, while other doses caused a decrease in semen motility and viability (Table 2).

Another parameter of the study was to determine of the integrity of membrane frozen-thawed. Based on the data on Table 2 in this study, fish oil had no significant effect on membrane integrity.

#### 4. Discussion

AI programs in large livestock including Kalang swamp buffaloes generally use frozen semen. There are various benefits of using frozen semen such as easy transportation, kept in long term, and efficient use of bulls to increase the potential of gene. Success of the semen freezing program can faciliate the improvement of swamp buffaloes population, for sperm can be kept for a long time and can be used as superior genetic stock. Sperm motility is a significant parameter in assessing the quality of spermatozoa, as it will affect the success of fertilization. Therefore, it is required to increase the high motility in post-thawing semen.

In this study, the results revealed that addition of concentration 150 mg fish oil supplementation in extender remarkably enhanced sperm motility before freezing. This significant increment in postthawed motility at dose 150 mg fish oil supplementation may be related with the collective effect of chemical structure of fish oil. Thus, the presence of concentration in the extender before freezing has provided better protection to the sperm motility or viability against cryopreservation-induced damages.

Sperm qualities including motility can be protected by fish oil during freezing process, because it is related to the content of ingredients from fish oil such as eicosapentaenoic acid, docosahexaenoic acid, saturated fatty acid, and PUFA. Guthrie and Welch[18] explained that the freezing process of semen can cause physical damage and disfunction due to excessive generation of reactive oxygen species. Furthermore, Nair *et al*[19] and Andrabi[8] revealed that pre-dominance of PUFA in sperm plasma membrane instigates the lipid peroxidation process which in turn reduces the semen qualities including sperm viability and motility. While, the part content fish oil of PUFA, eicosapentaneoic acid and docosahexaneoic acid allegedly play a role in sperm resistance to cooling and freezing procedures and may be correlated with the type of long chain PUFA content[20].

While, the previous study<sup>[14]</sup> conducted on bovine semen revealed a significant progress in post-thawing viability and sperm motility with the addition of 100 mg fish oil in the 100 mL skim milk-eggyolk extender. That is because one of the constituents of fish oil is PUFA with long chains and has been found in the semen of different species including man, bull and ram. These fatty acids could increase the fluidity of the membrane of plasma which is then accountable for improved struggle of the sperm to conditions of cold<sup>[21]</sup>. The membrane of plasma is the central location of injury prompted by cryopreservation that issues phospholipids into the nearby medium through of cold shock and becomes transiently leaky because of lipid phase transitions<sup>[22]</sup>.

In the study, as for the integrity of membrane of frozen-thawed semen, membrane integrity has not significant effect between control and all treatments after addition concentration of fish oil

#### Table 1

Effect of fish oil on viability, motility, abnormality and plasma membrane integrity before freezing in Kalang swamp buffalo bull semen.

			•	
Treatments	Viability	Motility	Abnormality	Plasma membrane integrity
0 (control)	60.25±0.51	58.25±0.51 <sup>b</sup>	17.50±2.52	61.51±1.96
50 mg fish oil	59.75±2.15	53.75±2.15 <sup>a</sup>	18.52±1.15	63.59±0.95
100 mg fish oil	60.39±0.15	$53.57 \pm 1.15^{a}$	17.25±0.56	62.25±1.28
150 mg fish oil	62.13±1.30	60.13±1.30 <sup>b</sup>	17.05±1.54	61.75±1.46
200 mg fish oil	61.26±2.10	50.26±3.10 <sup>a</sup>	18.75±1.55	62.15±1.70

Different superscripts along the column indicate the significant differences (P < 0.05) among groups (n=24).

#### Table 2

Effect of fish oil on viability, motility, abnorality and plasma membrane integrity of post-thawed Kalang swamp buffalo bull semen.

Treatments	Viability	Motility	Abnormality	Plasma membrane integrity
0 (control)	51.20±1.13 <sup>b</sup>	42.25±0.51 <sup>b</sup>	27.50±2.52	59.51±1.96
50 mg fish oil	43.23±1.52 <sup>a</sup>	$36.85 \pm 2.15^{a}$	25.52±1.07	58.98±0.95
100 mg fish oil	44.17±1.15 <sup>a</sup>	36.57±1.15 <sup>a</sup>	26.25±0.36	58.29±1.28
150 mg fish oil	52.17±2.41 <sup>b</sup>	43.13±1.30 <sup>b</sup>	23.05±1.24	57.05±1.46
200 mg fish oil	42.16±1.70 <sup>a</sup>	36.06±3.10 <sup>a</sup>	23.75±1.67	58.16±1.70

Different superscripts along the column indicate the significant differences (P < 0.05) among groups (n=24).

142

in the extender. These results are confirmed by previous reports by Abavisani *et al*[23] who revealed that the addition of PUFA straightly to extenders was not effective in protection of the sperm membrane.

In conclusion, addition of 150 mg in 100 mL skim milk-egg yolk extender can give a good result to the sperm motility of post-thawed Kalang swamp buffalo semen.

#### **Conflict of interest statement**

We declare that we have no conflict of interest.

#### Acknowledgments

We would like to thank the staff from the Central of Artificial Insemination of Banjarbaru, South Kalimantan Province, Indonesia.

#### References

- Triwidanningsih E, Praharani L. *Buffaloes in Indonesia*. Bogor, Indonesia: Research Institute for Animal Production; 2005, p.114-120.
- [2] Malik A, Laily M, Muhammad IZ. Effects of long term storage of seemen in liquid nitrogen on the viability, motility and abnormality of frozen thawed Frisian Holstein bull spermatozoa. *Asian Pac J Reprod* 2015; 4(1): 22-25.
- [3] Andrabi MH, Ansari MS, Ullah N, Anwar M, Mehmood A, Akhter S. Duck egg yolk in extender improves the freezability of buffalobull spermatozoa. *Anim Reprod Sci* 2008; **104**: 427-433.
- [4] Qaisar S, Muhammad UM, Hamayun K, Asmaul H, Saima Q, Asima A, et al. Royal jelly supplementation in semen extender enhances post-thaw quality and fertility of Nili-Ravi buffalo bull sperm. *Anim Reprod Sci* 2016; 167: 83-88.
- [5] Pradeep K, Dharmendra K, Sikka P, Singh P. Sericin supplementation improves semen freezability of buffalo bulls by minimizing oxidative stress duringcryopreservation. *Anim Reprod Sci* 2015; **152**: 26-31.
- [6] Mohamed AE, Abd Elmonem M, Mamdouh H, Ashraf El, Magdy B, Abd Elraouf H, Ahmed B, Samy MZ. The effects of cholesterol loaded cyclodextrins on post-thawed quality of buffalo semen in relation to sperm DNA damage and ultrastructure. *Reprod Biol* 2017; **17**: 42-50.
- [7] Kumaresan A, Ansari MR, Abhishek G. Modulation of post-thawed sperm functions with oviductal proteins in buffaloes. *Anim Reprod Sci* 2005; 90: 73-84.
- [8] Andrabi SMH. Factors affecting the quality of cryopreserved buffalo (Bubalus bubalis) bull spermatozoa. Reprod Dom Anim 2009; 44: 552-569.
- [9] Bansal AK, Bilaspuri GS. Impacts of oxidative stress and antioxidants on semen functions. *Vet Med Int* 2011; 6: 861-867.

- [10]Stoeckel K, Nielsen LH, Fuhrmann H, Bachmann L. Fatty acid patterns of dog erythrocyte membranes after feeding of a fish-oil based DHA-rich supplement with a base diet low in n-3 fatty acids versus adiet containing added n-3 fatty acids. *Acta Vet Scand* 2011; 6: 53-57.
- [11]Nichols PD, Glencross B, Petrie JR, Singh SP. Readily available sources of long-chain omega-3 oils: Is farmed Australian seafood a better source of the good oil than wild-caught seafood. *Nutrients* 2014; 6: 1063-1079.
- [12]Hossain MS, Tareq K, Hammano KI, Tsujii H. Effect of fatty acids on boar sperm motility, viability and acrosome reaction. *Reprod Med Biol* 2007; 6(4): 235-239.
- [13]Kiernan M, Fahey A, Fair S. The effect of the *in vitro* supplementation of exogenous long-chain fatty acids on bovine sperm cell function. *Reprod Fertil Dev* 2013; 25(6): 947-954.
- [14]Malik A, Syarifdjaya M, Gunawan A, Erlina S, Jaelani A, Wibowo DB. Effect of fish oil addition to the skim milk-egg yolk extender on the quality of frozen-thawed Bali bull spermatozoa. *Kafkas Univ Vet Fak Derg* 2017; 23(4): 651-654.
- [15]Khumran AM, Yimer N, Rosnina Y, Ariff MO, Wahid H, Kaka A, et al. Butylated hydroxytoluene can reduce oxidative stress and improve quality of frozen-thawed bull semen processed in lecithin and egg yolk based extenders. *Anim Reprod Sci* 2015; **163**: 128-134.
- [16]Memon AA, Wahid H, Rosnina Y, Goh YM, Ebrahimi M, Nadia F. Effect of antioxidants on post-thaw microscopic, oxidative stress parameter and fertility of Boer goat spermatozoa in Tris egg yolk glycerol extender. *Anim Reprod Sci* 2012; **136**(1): 55-60.
- [17]Kaka A, Wahid H, Rosnina Y, Yimer N, Khumran AM, Behan AA, Ebrahimi M. Alpha-linolenic acid supplementation in Tris extender can improve frozen-thawed bull semen quality. *Reprod Domest Anim* 2015; 50(1): 29-33.
- [18]Guthrie HD, Welch GR. Determination of intracellular reactiveoxygen species and high mitochondrial membrane potential inpercoll-treated viable boar sperm using fluorescence-activated flowcytometry. J Anim Sci 2006; 84: 2089-2100.
- [19]Nair KS, RA Rizza, Brien PO. DHEA in elderly women and DHE or testosteron on elderly men. N Engl J Med 2006; 355: 1647-1659.
- [20]Castellano CA, Audet I, Bailey JL, Laforest JP, Matte JJ. Dietary omega 3 fatty acids (fish oils) have limited effects on boar semen stored at 17 °C or cryopreserved. *Theriogenology* 2010; **74**: 1482-1490.
- [21]Wathes DC, Abayasekara DR, Aitken RJ. Polyunsaturated fatty acids in male and female reproduction. *Biol Reprod* 2007; 77(2): 190-201.
- [22]Behera S, Harshan HM, Bhai KL, Ghosh KNA. Effect of cholesterol supplementation on cryosurvival of goat spermatozoa. *Vet World* 2015; 8(12): 1386-1391.
- [23] Abavisani A, Arshami J, Naserian AA, Sheikholeslami KMA, Azizzadeh M. Quality of bovine chilled or frozen-thawed semen after addition of omega 3 fatty acids supplementation to extender. *Int J Fertil Steril* 2013; 7(3): 161-168.